

Decoding Amyotrophic Lateral Sclerosis: A Systems Biology Approach

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ABSTRACT

Amyotrophic lateral sclerosis (ALS) is a neurodegenerative disorder characterized by the loss of upper and lower motor neurons in the motor cortex, brain stem, and the anterior horn of the spinal cord. The majority of ALS cases are classified as sporadic (sALS). There is a growing concern regarding the increased incidence in the number of sporadic ALS cases across the world, projected to increase by almost 70% in the next two decades. The etiology of sporadic ALS is currently unknown; however, epidemiological studies point to possible exposure of environmental triggers, including trauma and infections as risk factors for the development of motor neuron pathology. On a pathological basis, protein misfolding with the accumulation of cytoplasmic inclusions of TDP-43 are regarded as the hallmark feature of ALS pathogenesis. The cellular mechanisms that lead to protein aggregation are not completely understood, but appear to involve defects in autophagy, an intracellular autodigestive process that degrades misfolded proteins like TDP-43. This review will be split into two portions: (1) discuss the evidence regarding how various environmental risk factors, such as infections agents and physical trauma, can lead to neuropathological changes by disrupting autophagy in ALS; (2) discuss potential treatment options in the management of each environmental factor previously discussed.

Keywords: ALS; Infectious; Autophagy; TBI; Neurodegeneration

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INTRODUCTION

Amyotrophic lateral sclerosis (ALS) is regarded as an invariably fatal neurodegenerative disease characterized by a progressive loss of motor neurons in the spinal cord, brainstem, and corticospinal tract that commonly presents with progressive asymmetric distal weakness and/or bulbar dysfunction. Respiratory failure typically limits survival to 2–5 years after disease onset, although the clinical presentation and course of ALS can be quite heterogeneous with a high degree of variability in the age at onset, the site of onset, and the rate of disease progression among different patients.^{1–4} In fact, clinical heterogeneity exists even between those with the same genetic familial form.⁵

In ALS, one of the primary misfolded proteins is TDP-43, a 414 amino acid-long nuclear RNA binding protein linked to familial and sporadic cases.⁶ TDP-43 mutations at the C-terminal increase its propensity to aggregate, and it has been shown to disinhibit viral expression, which in turn can provoke a heightened immune response characterized by pro-inflammatory cytokines such as transforming growth factor-beta (TGF- β).^{7,8} The connection between TDP-43 and RNA pathology from viral or enteropathogens was initially made based upon the discovery that TDP-43 inhibits HIV-1 gene expression by blocking the assembly of its transcription complexes.⁹ A further clue that ALS can be precipitated by RNA viruses is based upon the fact that several pathogens exhibit tropism for anterior horn cells, including enteroviruses, coxsackievirus, echovirus, polio, HIV, HTLV-1, and HERV-K.¹⁰ As mentioned, retrovirus activity and other microbial pathogens can elicit the production of pro-inflammatory cytokines such as TGF- β , which in turn induces membrane fibrosis leading to vascular remodeling, impaired perivascular cerebrospinal fluid (CSF) distribution, reduced compliance of glymphatic vessels, and defects in autophagy.¹¹ Autophagy is a multi-step pathway for intracellular and extracellular processes that are essential to degrade pathological protein aggregates and removal through the glymphatic system. The glymphatic system is a glial cell-mediated lymphatic system of the brain that can be disrupted by several environmental

factors. From an intracellular perspective, functional lysosomes digest misfolded proteins through pH-dependent proteolytic enzymes. However, if lysosomes are downregulated, extracellular clearance is required to remove macromolecules from the interstitial fluid via the CSF and glymphatic system. Environmental factors, notably sleep disturbances, infections, and repetitive head injuries, all have been proposed as risk factors for the development of ALS. This review will discuss the pathophysiology of ALS and propose addressing the neurodegenerative disease utilizing a systems biology approach. A systematic approach to ALS offers a new paradigm that addresses the interdependence of the central nervous system, gastrointestinal system, and the immune system as evidence suggests that a multimodal approach rather than a single intervention may have greater effectiveness.

PART 1

TDP-43, RHO KINASES, AND INFECTIONS

TDP-43 a nuclear RNA binding protein, is intimately involved in the regulation of autophagy, an essential process that allows cells to maintain proteostasis by recognizing and degrading pathological proteins, whereas mutations in TDP-43 result in protein aggregation. One of the pathways involved in autophagy is mediated through Rho kinase signaling. Excessive Rho activation provokes neurite retraction and neuronal apoptosis, which may be linked to the pathogenesis of ALS.^{12,13} Rho kinases have been proposed to be intimately involved in ALS as *in vivo* studies utilizing pathway enrichment analysis have demonstrated significant enrichment of Rho GTPases in late-stage ALS.¹⁴ The connection between TDP-43 and Rho activity is supported by studies where the cellular knockdown of normal TDP-43 function induces cell death through the disruption of the Rho family of GTPases, resulting in reduced autophagy and the misfolding/aggregation of RNA proteins.¹⁵

The Rho family of GTPases act as master transcription termination factors and have the ability to regulate neuronal morphology and survival by providing stability for microtubules in motor neurons.^{16,17} Rho proteins are the targets of various bacterial enteropathogens, including toxicogenic *Clostridium* species and the cytotoxic necrotizing factors from *Escherichia coli*, which may exert their toxicity through inhibiting autophagy by altering GTP-binding proteins.^{18,19} It has been demonstrated that inhibition of the Rho GTPase family of cytoskeletal regulators causes a reduction in viral infection.²⁰ Several RNA viruses, as well as enteropathogens, induce cytoskeletal abnormalities that target motor neurons through the activation of Rho-GTPases. In HIV infection, for example, the virus causes activation of the Rho GTPase family of cytoskeletal regulators within the host CD4⁺ T cells, leading to optimal infectivity.²⁰ Furthermore, viral pathogens like HIV can spread through the enhancement of Rho-dependent migration of monocytes and macrophages.²¹

Additionally, the cytotoxicity of fungal species, including *Aspergillus* and *Fusarium* species, involves disrupting the Rho GTPase signaling cascade through the release of gliotoxins that alter Rho signaling to favor their infectivity.^{22,23} *Aspergillus* infection leads to the formation of actin aggregates through disruption of RNA signaling, which in turn inhibits the assembly of v-ATPase on lysosomal membranes.²⁴ Exposure of neurons to fungal neurotoxins elicits a significant increase in abnormal TDP-43 translocation from the nucleus to the cytoplasm, which furthers the supportive link between infections and ALS pathogenesis.²⁵ Evidence for the potential connection between fungal exposure and motor neuron disease is supported as fungal antigens and DNA have been detected in CSF and brain tissue from ALS patients using PCR analysis.²⁶ In summary, disruption of TDP-43 pathways involving Rho kinases may be secondary to infection-mediated toxins that disrupt microtubules.

INFECTION PATHOGENESIS

It is well established, for instance, that motor neuron pathology can be precipitated by enteropathogenic viruses that exhibit tropism for anterior

horn cells, such as poliovirus. A growing list of RNA viruses, bacterial, and fungal species are capable of remaining dormant in the nervous system by eluding host immune defense mechanisms by inhibiting lysosomal-mediated protein degradation. Lysosomes promote autophagy and are an essential element of the innate and adaptive immune system for antigen processing.^{27,28} Multiple intracellular organisms survive within their hosts based upon their ability to impair lysosomal-based enzymes responsible for degrading pathological proteins formed by infectious particles.

It has been proposed that many intracellular pathogens resist sterile eradication based upon their ability to inhibit lysosomal vacuolar ATPase.^{29,30} Thus, the connection between ALS and various microbial toxins is linked to the disruption of host vacuolar-type ATPase (v-ATPase), leading to reduced lysosomal-mediated autophagy.³¹ Indeed, loss of v-ATPase-mediated lysosomal acidification has been proposed as a primary mechanism in ALS pathogenesis.³² Further support for an infectious etiology in ALS is based upon the unexplained findings of defects in iron homeostasis in patients. In several studies examined by meta-analysis, serum iron levels were shown to be lower and ferritin levels are higher in ALS patients.³³ Iron deficiency may result as a consequence of enteropathogens that divert dietary iron from their host, whereby microbial infections hijack iron delivery from hosts to promote their survival.^{34,35} *E. coli*, for instance, acquires enhanced virulence through overexpression of several iron acquisition systems leading to host iron deficiency.³⁶

THE ROLE OF IRON IN PATHOGENESIS

Lysosomes, as the intracellular site of autophagy, are involved in both innate and adaptive immune functions, including foreign material recognition (bacterial, parasitic, and viral) and antigen processing.²⁸ Multiple intracellular organisms survive within the lysosome based upon parasitic-mediated deacidification mechanisms resulting in defective xenophagy – a form of autophagy where host cells ingest foreign antigens – by interrupting lysosomal vacuolar ATPase and its ability to promote lysosomal acidification.^{29,30}

The chronicity of infectious diseases occurs by their ability to hijack RNA metabolism and thereby interrupt vacuolar inhibit lysosomal vacuolar ATPase and its ability to promote lysosomal acidification.^{29,30} In ALS, it is feasible that host Rho proteins are the targets of various bacterial and viral toxins that collude to cause cell death by inhibiting the assembly of host v-ATPase, disrupting lysosomal activity and autophagy.³¹ Indeed, loss of v-ATPase-mediated lysosomal acidification has been proposed as a primary mechanism in ALS pathogenesis.³²

Lysosomes are reservoirs of reactive ferrous iron that possess a system for cation transport through endosomal Fe(2+) transporter that coordinates iron metabolism, where it functions to transfer the endosomal free Fe(2+) into the cytoplasm.³⁷ Unbiased genetic screens discovered that lysosomal acidity was maintained via iron homeostasis.³⁸ Iron deficiency in ALS patients can lead to lysosomal v-ATPase inhibition and defective xenophagy, and the maintenance of a highly acidic pH is dependent upon iron homeostasis, which is essential for regulating many functions of lysosomes, including the activity of the v-ATPase to maintain autophagy.^{39–41} It is worth noting that defects in v-ATPase activity and alkalinized lysosomes have been reported in ALS.³²

Iron deficiency results in the up-regulation of divalent metal transporter 1 (DMT1), an iron-transport protein that contributes to antimicrobial function.³⁵ The gene that encodes DMT1 iron transport has been reported to result in a faster rate of progression than in ALS patients who do not harbor the same gene defect.⁴² It is interesting to note that DMT1 normally acts to confer resistance against bacterial infection by regulating proton/metal-ion cotransport in lysosomes.⁴³ However, it was recently demonstrated that hepatitis C and other viral pathogens may inhibit the expression of DMT1 to redirect iron towards viral transcription.⁴⁴ Thus, the virulence of chronic intracellular infections is facilitated by interrupting host post-transcriptional regulation of iron transport through DMT1.⁴⁵ Lysosomal dysregulation through alkylation and dysfunctional DMT1 are major pathogenic factors that contribute to the accumulation of protein aggregates and may represent the root cause of ALS pathogenesis.

INFECTION-MEDIATED ALTERED IMMUNE HOMEOSTASIS

Loss of immune homeostasis and/or early deregulation of immune responses may represent one of the key initial steps in disease pathogenesis in ALS, leading to either excessive inflammation and/or an inefficient immune response.⁴⁶ For example, ALS progression has recently been intricately linked to patient T cell profiles. T regulatory cells (Tregs) that balance the activation/inhibition of the immune system's response to infection negatively correlate to disease severity, that is, low quantity are predictive for rapid disease progression and short survival.⁴⁷ Further, T cell subtypes Th1 and Th17 presented a shift towards cytokine production, including interferon-gamma, whereas anti-inflammatory Th2 and T regulatory cells were decreased. Pro-inflammatory serum cytokines include interleukin (IL)-1 β , and IL-6.⁴⁸ In addition to declining Treg activity, TGF- β 1 – an anti-inflammatory cytokine – has been reported as a potential marker of disease ALS severity as there is a strong negative correlation between the cytokine and ALS function.⁴⁹ Elevations of TGF- β levels are often found in association with chronic infections as TGF- β is produced by the infected intestinal epithelium, including *Helicobacter pylori*, and hepatitis B and C virus.^{50–53}

Despite its typically known anti-inflammatory activity, it is interesting to note that elevated levels of TGF- β can adversely affect the lymphatic system by inducing fibrosis in the perivascular network of blood vessels, leading to impaired perivascular CSF distribution and reduced compliance of lymphatic vessels.¹¹ In clinical studies of ALS, evidence for the involvement of TGF- β in ALS pathogenesis includes findings of enhanced fibrosis and TGF- β 1 immunoreactivity in ALS patients greater than twofold over controls.⁵⁴ Evidence of systemic fibrosis has included myocardial imaging demonstrating increased magnetic resonance imaging (MRI) T1 enhancement in ~80% of ALS patients compared to ~25% of controls with a trend of myocardial fibrosis observed in ~25% of the patients and ~10% of controls.⁵⁵

VASCULAR DYSFUNCTION

In a similar manner as chronic infections, bacterial infections, including periodontal bacteria, gut

microbiota, *Helicobacter pylori*, and *Chlamydia* are linked to the pathogenesis of atherosclerosis through immunological cross-reactions.⁵⁶ Thus, ALS may involve infection-mediated changes in the vascular system as well. Evidence for vascular dysfunction in ALS has been supported by a number of studies, including peripheral arterial tonometry, which revealed significant declines of peripheral arterial endothelial functions in ALS, a marker of vascular dysfunction.⁵⁷ Microvascular disturbances involving the degeneration of endothelial cells surrounding the blood–brain barrier and in the ventral horn of patients have also been reported, demonstrating the role of vascular degeneration in ALS pathogenesis.⁵⁸

TDP-43 has been shown to result in reduced blood circulation and alterations in vascular function.⁵⁹ The protein deposits have been found both as cytoplasmic inclusions and in carotid and cerebrovascular plaques.⁶⁰ This strongly suggests that ALS not only involves TDP-43-mediated alterations in motor neurons but involves the vascular system as well. Furthermore, TDP-43(+) inclusions reduce blood circulation and may cause aberrant vasculogenesis in association with motor neuron pathology.⁵⁹

TRAUMA-MEDIATED GLYMPHATIC DYSFUNCTION

As mentioned previously, head injuries pose a risk factor for sporadic ALS. The connection between traumatic brain and spine injuries and neurodegeneration is supported by strong epidemiological evidence of repetitive head injury as an independent risk factor for the development of sporadic ALS.⁶¹ Moreover, consortium studies found that two or more head injuries are associated with a greater than threefold increased risk of ALS.⁶² Head and neck trauma may lead to disruption of the blood–brain barrier in tandem with ischemic changes and vascular hypoperfusion of motor neurons. In autopsied samples of the cervical spinal cord from ALS patients, perivascular hemoglobin deposits with the accumulation of fibrin and thrombin were found in the majority of patients.⁶³ Furthermore, TDP-43 levels are differentially affected by the number and magnitude of blast exposures, increasing following a greater number of exposures at various intensities.⁶⁴

At the spinal cord level, ischemic changes may be precipitated by dural displacement and epidural venous swelling in the spinal cord. For instance, in Hirayama disease, a disorder in the differential of ALS, results in ischemic myelopathy due to displacement of the dura, and perivenular compression is best visualized by dynamic flexion MRI studies.⁶⁵ Support for structural impediments in motor neuron disease includes recent MRI findings that have confirmed cervical spine abnormalities in patients with ALS and reduced CSF flow.⁶⁶

The glymphatic system refers to a network of venous perivascular spaces that directs interstitial fluid into the CSF.⁶⁷ The glymphatic system extends longitudinally down the entire length of the spinal canal through perivertebral veins that can become compressed due to neck trauma. Support for glymphatic failure in neurodegeneration has been demonstrated in Alzheimer disease (AD), where amyloid deposits were found in high concentrations in the cervical and axillary lymph nodes, which are part of the glymphatic system.⁶⁸ Further support for glymphatic failure in neurodegenerative disease was the recent finding that efflux of CSF containing amyloid- β and phosphor-tau has been reduced in patients with AD, compared to age-matched controls.⁶⁹ Parkinson disease has also been associated with a sustained reduction of CSF-glymphatic fluid transport.⁷⁰

Comparison of ALS and control groups revealed a reduction in CSF flow magnitude and increased flow propagation velocities in the ALS cohort, with the ALS cohort displaying nearly zero CSF flow along the entire spinal canal, supporting the notion that glymphatic dysfunction is directly linked to ALS pathogenesis.⁷¹ Recent findings suggest that glymphatic dysfunction may be the root cause of autophagic defects after traumatic head injuries, resulting in neurodegeneration due to insufficient venous drainage in the CNS.⁷²

Trauma can also induce damages to the autonomic nervous system. In ALS, degeneration of the central autonomic network, including the hypothalamus and the vagal nerve, is disrupted as evidenced by ultrasound-based diagnostics.⁷³ Impaired cardiovagal function can lead to an unexpected death in patients as autonomic dysfunction is often found in patients with neurodegenerative diseases. In summary, in a

similar manner as intracellular infections can precipitate neurodegenerative changes based upon defects in intracellular autophagy, traumatic events involving the head and neck can reduce autophagy through biophysical disruption of the glymphatic system.

PART 2

ADDRESSING INFECTION AND MITOCHONDRIAL DYSFUNCTION

As mentioned above, there is both circumstantial and direct evidence of a potential association with chronic intracellular pathogens. While the theory of ALS as an infectious disease is still in its infancy, RNA-based viral, bacterial, and even fungal pathogens have been shown to disrupt microtubules leading to motor neuron pathology. Recent findings in models of ALS have demonstrated putative benefits in inhibiting TDP-43 protein aggregates.⁷⁴

Doxycycline (a tetracyclic antibiotic) and its derivatives have been shown to have neuroprotective effects by inhibiting amyloid aggregation and also inhibiting aggregation of α -synuclein, the pathological protein in Parkinson disease.⁷⁴ In ALS mouse models, doxycycline acts to suppress the expression of human TDP-43 and inhibits motor neuron loss. The administration of doxycycline showed a significant increase in muscle innervation, the rescue of motor impairments, and a dramatic extension of lifespan, even after neurodegeneration and motor dysfunction.⁷⁵ Interestingly, doxycycline also decreases RhoA-GTPase activity, which may be part of its neuroprotective mechanism.⁷⁶ Sub-antibiotic doses of doxycycline also appear to be high enough to interfere with the production of α -synuclein toxic species, the pathological aggregate in Parkinson disease pathogenesis.⁷⁷ However, multiple bacterial organisms have developed different mechanisms of resistance mediated by drug efflux mechanisms involving the TetA protein (tetracycline activating protein), which have been identified in the human gut metagenome that confers resistance to tetracycline.^{78,79}

Resistance to doxycycline may potentially be overcome by natural monoterpenoids such as β -thujaplicin (hinokitiol). A comprehensive study

found that the natural plant alkaloid overcame TetA tetracycline bacterial resistance to doxycycline.⁸⁰ On its own, β -thujaplicin possesses high antimicrobial activity against various oral, nasal, and nasopharyngeal pathogens, including *Streptococcus mutans*, *Porphyromonas gingivalis*, and prevents biofilm formation in both antifungal-susceptible and antifungal-resistant strains of *Candida* species.^{81,82} β -Thujaplicin has also been shown to protect against prion-induced neurotoxicity by inducing autophagy via the activation of AMPK, which in turn increases the formation of functional autolysosomes to promote autophagy.^{83,84} Furthermore, β -thujaplicin binds to both ferrous and ferric iron with high affinity and maintains iron homeostasis, which can restore iron transport into, within, and/or out of cells.⁸⁵

Nonpharmacologically, taurine – a semi-essential amino acid – induces the production of CD4⁺CD25⁺FoxP3⁺ Treg cells, which may restore the population of Treg cells and normalize inflammatory responses.⁸⁶ Taurine also has demonstrated an anti-fibrotic effect in several animal models where experimentally induced elevations of TGF- β 1 were significantly suppressed by taurine and that effectively reduced the abnormal deposition of collagen.⁸⁷ In ALS, the immunoreactivity of the taurine transporter in spinal cord motor neurons of ALS transgenic mice and in spinal motor neurons of patients with ALS has been found to be increased.⁸⁸ Taurine also has inherent antimicrobial effects in addition to its anti-inflammatory effects. Neuroinflammation is a host response to infections where neutrophils are recruited and undergo an oxidative burst to diminish microbial burden. Taurine reacts with hypochlorous acid (HOCl), an antibacterial pro-oxidant produced by myeloperoxidase, resulting in the generation of taurine chloramine (TauCl), a weak oxidant with mild cytotoxicity.⁸⁹ Intriguingly, *N*-acetylhomotaurine (a pharmaceutical analog of taurine approved by the FDA for alcohol cessation) was shown to inhibit the accumulation of TDP-43 protein in the cytoplasm when combined with baclofen, a muscle-relaxing agent.⁹⁰ This effect may be attributed in part to homotaurine's ability to indirectly inhibit *N*-methyl-D-aspartic acid receptors, a primary pathological target of Riluzole, a drug approved by the FDA for ALS.⁹¹

Riboflavin is a water-soluble B vitamin that is involved in a diffuse array of metabolic processes

involving the biosynthesis of coenzyme A, coQ10, heme, and folic acids to support methylation pathways.⁹² Riboflavin, perhaps by supporting mitochondrial activity, plays a role in elevating heat shock proteins as an inherent defense against microbial challenges by acting as an agonist of MHC-related protein-1, which in turn facilitates mucosal-associated invariant T cells.⁹³ Motor neurons are selectively impaired during riboflavin deficiency states by which hypomagnesemia along with low magnesium may impede ATP production. In case studies, riboflavin at 150 mg/day improved muscle strength and reduced plasma lactate and alanine concentrations in a patient with mitochondrial complex I deficiency.^{94,95} To illustrate its potential, autosomal recessive mutations in the riboflavin transporter genes cause a condition referred to as Brown–Vialletto–Van Laere (BVVL), a rare hereditary form of ALS. Reduced autophagic flux in motor neurons may be partially rescued by riboflavin supplementation.⁹⁶ A case report of juvenile ALS – phenotype secondary to a genetic defect in riboflavin transporter deficiency – was dramatically improved after high-dose riboflavin replacement therapy.⁹⁷ It is interesting to note that riboflavin has been shown to reduce the virulence of *E. coli* and *S. aureus*, and the production of pro-inflammatory cytokines induced by lipopolysaccharide.^{93,98} Analogues of the riboflavin biosynthesis pathway, such as roseoflavin, have been shown to exert bactericidal effects by targeting and downregulating the riboflavin riboswitch in bacteria.⁹⁹

Carnitine also plays an important role in energy metabolism and has been shown to improve inspiratory capacity.^{100–102} A case report found that L-carnitine and riboflavin improved clinical symptoms and normalized complex I activity in a 6-year-old with myopathy (associated with complex I deficiency) and pure motor neuropathy.¹⁰³ After supplementation, the patient improved, and by 18 months only slight weakness in the peroneal muscles was detected on clinical exam. While the combination of magnesium, taurine, riboflavin, and carnitine have not been previously conceived as yet for a clinical trial for ALS, its purported mechanisms of action seem worth considering to assess in patients with evidence of mitochondrial dysfunction, while also addressing the infection basis of disease pathology.

ADDRESSING ALS AS A VASCULAR DISEASE

RhoA/Rho-kinase pathways have been demonstrated to play a role in the pathogenesis of vasospasm, glaucoma, arteriosclerosis, ischemic injury, and hypertension.¹⁰⁴ Fasudil, a Rho kinase inhibitor, relaxes smooth muscle and is approved for the treatment of cerebral vasospasms in Japan.^{105–108} Rho-kinase inhibitors are now approved for use in glaucoma by lowering intraocular pressure (IOP) and enhancing aqueous humor outflow.^{109,110} There is a potential association between glaucoma and ALS pathology due to common mutations in optineurin, which plays an essential role as an autophagy inducer.^{111–113} These findings led to an ALS study whereby fasudil showed improvement in respiratory function in a small cohort of patients that was followed by a current phase 2 multicenter, double-blind, randomized, placebo-controlled trial of fasudil in ALS patients.¹¹⁴

Berberine is a natural isoquinoline alkaloid that acts in part as a Rho kinase inhibitor and affects sigma receptor 1, similar to many antidepressant drugs.¹¹⁵ Sulfate salts of berberine readily cross the blood–brain barrier and have been used clinically for their antimicrobial and antifibrotic effects, and promote the integrity of the blood–brain barrier by inhibiting Rho and activating AMPK to promote autophagy.^{116,117} In ALS, berberine has been shown to reverse the processing of insoluble TDP-43 aggregates through the activation of the autophagic degradation pathway by acting as a Rho kinase inhibitor.¹¹⁸ Berberine derivatives promote degradation of mutant proteins of ALS in spinal and bulbar muscular atrophy, and has overlapping putative effects on misfolded proteins in amyotrophic lateral sclerosis and frontotemporal-lobar degeneration.¹¹⁹

In regards to lipid-based risk factors in ALS, analyses have demonstrated that higher low-density lipoprotein (LDL) levels are causally associated with an increased risk of developing ALS in both European and East Asian populations.¹²⁰ While there is caution in prescribing statins for patients with myopathies, it is interesting to note that cholestyramine binds bile acids and reduces LDL cholesterol, and is approved by the FDA for the management of dyslipidemias. Interestingly, regarding its potential

use in ALS, cholestyramine binds mold toxins produced by various fungal species.¹²¹

ADDRESSING PHYSICAL TRAUMA

Biophysical methods, including the use of focused ultrasound (FUS) modalities, have been shown to improve glymphatic dysfunction. For example, it was recently shown that focused ultrasound was able to reduce the amyloid burden and improve cognition in a small cohort of AD patients. Therapeutic FUS has been shown to reduce A β plaque in a small group of patients with AD.¹²² Mechanistically, FUS has been shown to expand the perivascular spaces of the brain to enhance the dispersion of protein aggregates and induce endothelial production of nitric oxide (NO) release to augment microvascular perfusion.¹²³ Potential therapeutics that can improve glymphatic function include assessment and correction of abnormalities through manual therapies, such as atlas orthogonal modalities treatment to relieve chronic cerebrospinal venous insufficiency secondary to mechanical compression of the epidural space.¹²⁴

ADDRESSING RESPIRATORY DYSFUNCTION

Key respiratory muscle dysfunction in ALS involves the diaphragm, accessory muscles of respiration, and bulbar muscles, which can lead to aspiration, diminished airway clearance, ineffective cough, recurrent pulmonary infections, and chronic hypercapnic respiratory failure.^{125–127} Clenbuterol, a beta-agonist, is currently used in trials for ALS, but common side effects, including tremor and headache and dose-dependent abnormalities of laboratory values including hypokalemia and hypoglycemia, may preclude its long term use.¹²⁸ Respiratory muscle dysfunction in ALS can be addressed by pharmacological agents such as aminophylline, an FDA-approved drug for the treatment of chronic obstructive pulmonary disease, by acting as an adenosine A2a receptor antagonist.¹²⁹ Aminophylline induces long-lasting phrenic motor facilitation and may provide an effective therapeutic strategy in the treatment of patients with ventilatory control disorders, such as obstructive sleep apnea, or respiratory insufficiency after a spinal injury or during neurodegenerative diseases.¹³⁰

In a randomized controlled study in ALS, the administration of aminophylline led to improvements in forced vital capacity, maximal mouth inspiratory and expiratory pressures ($P_{I_{max}}$ / $P_{E_{max}}$), and handgrip strength.¹³¹

ADDRESSING PSYCHOLOGICAL FACTORS AND IMPROVING SLEEP

There are anecdotal reports that lithium is helpful in patients with pseudobulbar palsy, a common co-morbid feature in ALS patients characterized by emotional lability.¹³² Mechanistically, a further rationale to consider lithium on an empirical basis is its putative effects on RNA metabolism in ALS. Lithium is known to act as an autophagy enhancer by inhibiting GSK-3 β and has been shown (in low doses) to reduce A β plaques and p-tau levels.¹³³ Lithium also has supportive effects in collapsin response mediator proteins (CRMPs) that act to stabilize microtubules via the enhancement of microtubule-associated protein 1 light chain 3 (LC3) to support autophagy.^{134–137} Mutations in CRMP interact with RhoA to suppress neurite outgrowth.¹³⁸ In ALS mouse models, genetic corrections to CRMP2 phosphorylation delayed the progression of motor symptoms in the mutant model.^{139,140} Lithium regulates the activity of CRMP by inhibiting GSK3 β expression, which is upregulated in ALS.¹⁴¹ Lithium administration also has been shown to promote autophagy flux to protect motor neurons in the spinal cord.^{142,143} Lithium has also been shown to result in a marked increase in LC3-positive vacuoles in mouse models and has shown therapeutic potential in the clinical setting for ALS and its mood disorder-related pathology.^{144–148}

Antidepressants, especially serotonin agonists, may have a protective effect in ALS by acting through sigma 1 receptor (and/or AQP4 receptors) expressed in motor neurons in the brainstem and in spinal motor neurons.¹⁴⁹ Activation of sigma 1 receptors is currently being investigated in ALS and Huntington disease, which through the restoration of autophagic flux may be neuroprotective.^{150,151} Recent studies have demonstrated that defects in SigR1 receptor binding are found in ALS patients.¹⁵² Sig-1R agonists have been found to increase Sig-1R expression and translocation to relieve blood–brain barrier dysfunction; meanwhile both Sig-1R knockout and Sig-1R

antagonists block the neuroprotective functions of the receptor.¹⁵³

Untreated depression can lead to anorexia and weight loss due to reduced caloric intake, and this reduction in consumption is an indicator of poor prognosis in ALS patients. Motor neurons require vast amounts of energy utilized by the mitochondria and reduced food intake and anorexia may exacerbate muscle catabolism.¹⁵⁴ As expected, caloric restriction shortened the lifespan of ALS mice models.¹⁵⁵ Conversely, performed with mutant SOD1^{G93A}, SOD1^{G86R}, and TDP43^{A315T} overexpressing mice, a high-fat diet delayed disease onset and extended survival, and in other studies attenuated motor neuron loss.^{156,157} In a double-blind, placebo-controlled trial, a high-fat hypercaloric diet led to a slower decline in ALSFRS-R scores compared to matched controls.¹⁵⁸

Pain is often an under-recognized symptom both in depression and in ALS patients. Palmitoylethanolamide (PEA), an endogenous compound belonging to the family of *N*-acylethanolamines, has been shown to alleviate pain signals by acting as a cannabinoid CB2 receptor agonist, also demonstrating beneficial effects in a small pilot study for depression.^{159,160} PEA also inhibits the Ca²⁺-dependent release of glutamate, and as excessive glutamate has been associated with many neurodegenerative diseases, PEA appears to be a potentially strong mechanistically based compound to consider in ALS patients.¹⁶¹ In this regard, PEA has been shown to reduce the rate of acetylcholine receptor desensitization at the neuromuscular junction in an average of 70% in ALS patients.¹⁶² Based upon these findings, PEA treatment was examined in a small pilot study of 28 ALS patients to receive 50 mg riluzole twice daily plus 600 mg PEA twice daily. No significant reductions in forced vital capacity were detected by pairwise comparisons during the analysis in PEA-treated patients.¹⁶²

Buspirone is an anxiolytic drug with robust serotonin receptor 1A (Htr1a) agonist activity that acts as a respiratory stimulant through brainstem chemoreflex afferents in central apnea.¹⁶³ The drug has also been shown to reduce hypocapnic central sleep apnea in chronic spinal cord injury patients, in which hypocapnic sleep apnea is a common complication in ALS.¹⁶⁴ Pharmacologically, buspirone not only acts as a serotonin agonist but also has an affinity for sigma

receptor 1 (SigR1), which plays an important role in regulating motor neuron function and autophagy – processes that are critical in ALS.¹⁶⁵ In randomized double-blind, placebo-controlled multicenter studies (phase III) with xaliproden – a 5-HT_{1A} receptor agonist – lead to improvements in vital capacity that are commonly reduced during sleep in ALS patients.¹⁶⁶

Ashwagandha extract has been shown to result in significant improvement in overall sleep compared to placebo in the dosage of ≥600 mg/day.¹⁶⁷ Regarding its potential putative effects in neurological and inflammatory disorders, withaferin A – bioactive constituent from *Withania somnifera* – has been shown to mitigate inflammatory cytokines, including nuclear factor kappa-B (NF-κB) and tumor necrosis factor-alpha (TNF-α), as well as diminish the expression of pro-fibrotic proteins.¹⁶⁸ Withaferin A also induces heat shock proteins that are instrumental in promoting protein folding.¹⁶⁹ Oral analogs of withaferin-A act as an antagonist of NF-κB and have been shown to reduce TDP-43 proteinopathy in the brain and spinal cord of transgenic mice expressing human TDP-43 mutations and restored the synthesis of neurofilament proteins.¹⁷⁰

Activation of melatonin MT₁ and MT₂ receptors exerts neuroprotective effects by inhibiting NF-κB activation, protecting neurons from lipopolysaccharide toxicity.¹⁷¹ In animal models of sepsis, melatonin administration inhibited the disruption of BBB permeability by attenuating microglial toll-like receptor 4/NF-κB signaling.¹⁷² Melatonin also plays a key role in autophagy by the regulation of autophagy mediated by the phosphorylation of AMPK.¹⁷³ Retrospective analysis regarding the impact of melatonin on progression and overall survival of ALS revealed that melatonin was shown to have a significantly decreased annualized hazard death rate, slowed the rate of functional decline, and change in predicted forced vital capacity score compared with the non-melatonin users.¹⁷⁴

CONCLUSION

In summary, ALS has recently been recognized as a systems biology disorder, given that neurogenerative changes occur not only in motor neurons but also in non-neurological tissue as well, including the

vascular system. This may account for significant co-morbidities related to sleep disorders, immune senescence, MRI changes in both white and gray matter, and pulmonary dysfunction. Both genetic and non-genetic factors contribute to the risk of developing ALS. Sporadic risk factors based upon epidemiological and anecdotal case studies point to a predilection for recurrent infections and head/neck trauma as possible contributing factors. These environmental insults converge on autophagy, an evolutionarily conserved process required to remove the pathological aggregates associated with ALS. Addressing ALS from a systems biology approach – including addressing sleep disorders, pain, musculoskeletal dysfunction, and breathing capacity – may yield unexpected synergistic effects in treatment beyond a single monotherapeutic agent.

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REFERENCES

1. Valbuena GN, Cantoni L, Tortarolo M, *et al.* Spinal cord metabolic signatures in models of fast- and slow-progressing SOD1G93A amyotrophic lateral sclerosis. *Front Neurosci.* 2019;13:1276. <https://doi.org/10.3389/fnins.2019.01276>.
2. Chiò A, Calvo A, Moglia C, *et al.* Phenotypic heterogeneity of amyotrophic lateral sclerosis: a population based study. *J Neurol Neurosurg Psychiatry.* 2011;82(7):740–6. <https://doi.org/10.1136/jnnp.2010.235952>.
3. Pupillo E, Messina P, Logroscino G, *et al.* Long-term survival in amyotrophic lateral sclerosis: a population-based study. *Ann Neurol.* 2014;75(2):287–97. <https://doi.org/10.1002/ana.24096>.
4. Fournier C, Glass JD. Modeling the course of amyotrophic lateral sclerosis [published correction appears in *Nat Biotechnol.* 2015;33(5):563]. *Nat Biotechnol.* 2015;33(1):45–7. <https://doi.org/10.1038/nbt.3118>.
5. Régál L, Vanopdenbosch L, Tilkin P, *et al.* The G93C mutation in superoxide dismutase 1: clinicopathologic phenotype and prognosis [published correction appears in *Arch Neurol.* 2006;63(7):963]. *Arch Neurol.* 2006;63(2):262–7. <https://doi.org/10.1001/archneur.63.2.262>.
6. Neumann M, Sampathu DM, Kwong LK, *et al.* Ubiquitinated TDP-43 in frontotemporal lobar degeneration and amyotrophic lateral sclerosis. *Science.* 2006;314(5796):130–3. <https://doi.org/10.1126/science.1134108>.
7. Johnson BS, Snead D, Lee JJ, *et al.* TDP-43 is intrinsically aggregation-prone, and amyotrophic lateral sclerosis-linked mutations accelerate aggregation and increase toxicity [published correction appears in *J Biol Chem.* 2009;284(37):25459]. *J Biol Chem.* 2009;284(30):20329–39. <https://doi.org/10.1074/jbc.M109.010264>.
8. Gonzalez D, Contreras O, Rebolledo DL, *et al.* ALS skeletal muscle shows enhanced TGF- β signaling, fibrosis and induction of fibro/adipogenic progenitor markers. *PLoS One.* 2017;12(5):e0177649. <https://doi.org/10.1371/journal.pone.0177649>.
9. Ou SH, Wu F, Harrich D, *et al.* Cloning and characterization of a novel cellular protein, TDP-43, that binds to human immunodeficiency virus type 1 TAR DNA sequence motifs. *J Virol.* 1995;69(6):3584–96. <https://doi.org/10.1128/JVI.69.6.3584-3596.1995>.
10. Alfahad T, Nath A. Retroviruses and amyotrophic lateral sclerosis. *Antiviral Res.* 2013;99(2):180–7. <https://doi.org/10.1016/j.antiviral.2013.05.006>.

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The authors declare they have no competing interests.

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J.L. collated all of the content in the manuscript. M.H. contributed to the preparation and revision of the manuscript. J.L. and M.H. contributed to the conception and interpretation of content, and are in agreement with all aspects of the work. All authors read and approved the final manuscript.

11. Howe MD, Furr JW, Munshi Y, *et al.* Transforming growth factor- β promotes basement membrane fibrosis, alters perivascular cerebrospinal fluid distribution, and worsens neurological recovery in the aged brain after stroke. *Geroscience*. 2019;41(5):543–59. <https://doi.org/10.1007/s11357-019-00118-7>.
12. Luo L. Rho GTPases in neuronal morphogenesis. *Nat Rev Neurosci*. 2000;1(3):173–80. <https://doi.org/10.1038/35044547>.
13. Stankiewicz TR, Pena C, Bouchard RJ, Linseman DA. Dysregulation of Rac or Rho elicits death of motor neurons and activation of these GTPases is altered in the G93A mutant hSOD1 mouse model of amyotrophic lateral sclerosis. *Neurobiol Dis*. 2020;136:104743. <https://doi.org/10.1016/j.nbd.2020.104743>.
14. Leoni E, Bremang M, Mitra V, *et al.* Combined tissue-fluid proteomics to unravel phenotypic variability in amyotrophic lateral Sclerosis. *Sci Rep*. 2019;9(1):4478. <https://doi.org/10.1038/s41598-019-40632-4>.
15. Iguchi Y, Katsuno M, Niwa JI, *et al.* TDP-43 depletion induces neuronal cell damage through dysregulation of Rho family GTPases. *J Biol Chem*. 2009;284(33):22059–66. <https://doi.org/10.1074/jbc.M109.012195>.
16. Linseman DA, Loucks FA. Diverse roles of Rho family GTPases in neuronal development, survival, and death. *Front Biosci*. 2008;13:657–76. <https://doi.org/10.2741/2710>.
17. Stankiewicz TR, Linseman DA. Rho family GTPases: key players in neuronal development, neuronal survival, and neurodegeneration. *Front Cell Neurosci*. 2014;8:314. <https://doi.org/10.3389/fncel.2014.00314>.
18. Popoff MR. Bacterial factors exploit eukaryotic Rho GTPase signaling cascades to promote invasion and proliferation within their host. *Small GTPases*. 2014;5. <https://doi.org/10.4161/sgtp.28209>.
19. Dufies O, Doye A, Courjon J, *et al.* *Escherichia coli* Rho GTPase-activating toxin CNF1 mediates NLRP3 inflammasome activation via p21-activated kinases-1/2 during bacteraemia in mice. *Nat Microbiol*. 2021;6(3):401–12. <https://doi.org/10.1038/s41564-020-00832-5>.
20. Lucera MB, Fleissner Z, Tabler CO, *et al.* HIV signaling through CD4 and CCR5 activates Rho family GTPases that are required for optimal infection of primary CD4+ T cells. *Retrovirology*. 2017;14(1):4. <https://doi.org/10.1186/s12977-017-0328-7>.
21. Drewry LL, Sibley LD. The hitchhiker's guide to parasite dissemination. *Cell Microbiol*. 2019;21(11):e13070. <https://doi.org/10.1111/cmi.13070>.
22. Latgé JP, Chamilos G. *Aspergillus fumigatus* and aspergillosis in 2019. *Clin Microbiol Rev*. 2019;33(1):e00140–18. <https://doi.org/10.1128/CMR.00140-18>.
23. Lahiani A, Yavin E, Lazarovici P. The molecular basis of toxins' interactions with intracellular signaling via discrete portals. *Toxins (Basel)*. 2017;9(3):107. <https://doi.org/10.3390/toxins9030107>.
24. Kogan TV, Jadoun J, Mittelman L, *et al.* Involvement of secreted *Aspergillus fumigatus* proteases in disruption of the actin fiber cytoskeleton and loss of focal adhesion sites in infected A549 lung pneumocytes. *J Infect Dis*. 2004;189(11):1965–73. <https://doi.org/10.1086/420850>.
25. French PW, Ludowyke R, Guillemin GJ. Fungal neurotoxins and sporadic amyotrophic lateral sclerosis. *Neurotox Res*. 2019;35(4):969–80. <https://doi.org/10.1007/s12640-018-9980-5>.
26. Alonso R, Pisa D, Marina AI, *et al.* Evidence for fungal infection in cerebrospinal fluid and brain tissue from patients with amyotrophic lateral sclerosis. *Int J Biol Sci*. 2015;11(5):546–58. <https://doi.org/10.7150/ijbs.11084>.
27. Levine B, Deretic V. Unveiling the roles of autophagy in innate and adaptive immunity. *Nat Rev Immunol*. 2007;7(10):767–77. <https://doi.org/10.1038/nri2161>.
28. Bonam SR, Wang F, Muller S. Lysosomes as a therapeutic target. *Nat Rev Drug Discov*. 2019;18(12):923–48. <https://doi.org/10.1038/s41573-019-0036-1>.
29. Huang J, Brummell JH. Bacteria-autophagy interplay: a battle for survival. *Nat Rev Microbiol*. 2014;12(2):101114. <https://doi.org/10.1038/nrmicro3160>.
30. Abdoli A, Alirezai M, Mehrbod P, Forouzanfar F. Autophagy: The multi-purpose bridge in viral infections and host cells. *Rev Med Virol*. 2018;28(4):e1973. <https://doi.org/10.1002/rmv.1973>.
31. Perez-Nadales E, Nogueira MF, Baldin C, *et al.* Fungal model systems and the elucidation of pathogenicity determinants. *Fungal Genet Biol*. 2014;70(100):42–67. <https://doi.org/10.1016/j.fgb.2014.06.011>.
32. Şentürk M, Mao D, Bellen HJ. Loss of proteins associated with amyotrophic lateral sclerosis affects lysosomal acidification via different routes. *Autophagy*. 2019;15(8):1467–9. <https://doi.org/10.1080/15548627.2019.1609863>.
33. Wang L, Li C, Chen X, *et al.* Abnormal serum iron-status indicator changes in amyotrophic lateral sclerosis (ALS) patients: a meta-analysis. *Front Neurol*. 2020;11:380. <https://doi.org/10.3389/fneur.2020.00380>.
34. Larson JA, Howie HL, So M. *Neisseria meningitidis* accelerates ferritin degradation in host epithelial cells to yield an essential iron source. *Mol Microbiol*. 2004;53(3):807–20. <https://doi.org/10.1111/j.1365-2958.2004.04169.x>.
35. Sousa Gerós A, Simmons A, Drakesmith H, *et al.* The battle for iron in enteric infections. *Immunology*. 2020;161(3):186–99. <https://doi.org/10.1111/imm.13236>.
36. Houamel D, Ducrot N, Lefebvre T, *et al.* Hcpidin as a major component of renal antibacterial defenses against uropathogenic *Escherichia coli*. *J Am Soc Nephrol*. 2016;27(3):835–46. <https://doi.org/10.1681/ASN.2014101035>.
37. MacKenzie EL, Iwasaki K, Tsuji Y. Intracellular iron transport and storage: from molecular mechanisms to health implications. *Antioxid Redox Signal*. 2008;10(6):997–1030. <https://doi.org/10.1089/ars.2007.1893>.

38. Weber RA, Yen FS, Nicholson SPV, *et al.* Maintaining iron homeostasis is the key role of lysosomal acidity for cell proliferation. *Mol Cell.* 2020;77(3):645–55.e7. <https://doi.org/10.1016/j.molcel.2020.01.003>.
39. Colacurcio DJ, Nixon RA. Disorders of lysosomal acidification—the emerging role of v-ATPase in aging and neurodegenerative disease. *Ageing Res Rev.* 2016;32:75–88. <https://doi.org/10.1016/j.arr.2016.05.004>.
40. Forgac M. Vacuolar ATPases: rotary proton pumps in physiology and pathophysiology. *Nat Rev Mol Cell Biol.* 2007;8(11):917–29. <https://doi.org/10.1038/nrm2272>.
41. Saroussi S, Nelson N. The little we know on the structure and machinery of V-ATPase. *J Exp Biol.* 2009;212(Pt 11):1604–10. <https://doi.org/10.1242/jeb.025866>.
42. Blasco H, Vourc'h P, Nadjar Y, *et al.* Association between divalent metal transport 1 encoding gene (SLC11A2) and disease duration in amyotrophic lateral sclerosis. *J Neurol Sci.* 2011;303(1–2):124–7. <https://doi.org/10.1016/j.jns.2010.12.018>.
43. Mackenzie B, Hediger MA. SLC11 family of H⁺-coupled metal-ion transporters NRAMP1 and DMT1. *Pflugers Arch.* 2004;447(5):571–9. <https://doi.org/10.1007/s00424-003-1141-9>.
44. Foka P, Dimitriadis A, Karamichali E, *et al.* HCV-induced immunometabolic crosstalk in a triple-cell co-culture model capable of simulating systemic iron homeostasis. *Cells.* 2021;10(9):2251. <https://doi.org/10.3390/cells10092251>.
45. Jiang S, Guo S, Li H, *et al.* Identification and functional verification of microRNA-16 family targeting intestinal divalent metal transporter 1 (DMT1). *Front Physiol.* 2019;10:819. <https://doi.org/10.3389/fphys.2019.00819>.
46. Béland LC, Markovinovic A, Jakovac H, *et al.* Immunity in amyotrophic lateral sclerosis: blurred lines between excessive inflammation and inefficient immune responses. *Brain Commun.* 2020;2(2):fcaa124. <https://doi.org/10.1093/braincomms/fcaa124>.
47. Beers DR, Zhao W, Wang J, *et al.* ALS patients' regulatory T lymphocytes are dysfunctional, and correlate with disease progression rate and severity. *JCI Insight.* 2017;2(5):e89530. <https://doi.org/10.1172/jci.insight.89530>.
48. Jin M, Günther R, Akgün K, *et al.* Peripheral pro-inflammatory Th1/Th17 immune cell shift is linked to disease severity in amyotrophic lateral sclerosis. *Sci Rep.* 2020;10(1):5941. <https://doi.org/10.1038/s41598-020-62756-8>.
49. Duque T, Gromicho M, Pronto-Laborinho AC, de Carvalho M. Transforming growth factor- β plasma levels and its role in amyotrophic lateral sclerosis. *Med Hypotheses.* 2020;139:109632. <https://doi.org/10.1016/j.mehy.2020.109632>.
50. Beswick EJ, Pinchuk IV, Earley RB, *et al.* Role of gastric epithelial cell-derived transforming growth factor beta in reduced CD4⁺ T cell proliferation and development of regulatory T cells during *Helicobacter pylori* infection. *Infect Immun.* 2011;79(7):2737–45. <https://doi.org/10.1128/IAI.01146-10>.
51. Alatrakchi N, Graham CS, van der Vliet HJ, *et al.* Hepatitis C virus (HCV)-specific CD4⁺ cells produce transforming growth factor beta that can suppress HCV-specific T-cell responses. *J Virol.* 2007;81(11):5882–92. <https://doi.org/10.1128/JVI.02202-06>.
52. Khorramdelazad H, Hassanshahi G, Nasiri Ahmadabadi B, Kazemi Arababadi M. High serum levels of TGF- β in Iranians with chronic HBV infection. *Hepat Mon.* 2012;12(11):e7581. <https://doi.org/10.5812/hepatmon.7581>.
53. Karimi-Googheri M, Daneshvar H, Nosratabadi R, *et al.* Important roles played by TGF- β in hepatitis B infection. *J Med Virol.* 2014;86(1):102–8. <https://doi.org/10.1002/jmv.23727>.
54. Si Y, Kim S, Cui X, *et al.* Transforming growth factor beta (TGF- β) is a muscle biomarker of disease progression in ALS and correlates with Smad expression. *PLoS One.* 2015;10(9):e0138425. <https://doi.org/10.1371/journal.pone.0138425>.
55. Rosenbohm A, Schmid B, Buckert D, *et al.* Cardiac findings in amyotrophic lateral sclerosis: a magnetic resonance imaging study. *Front Neurol.* 2017;8:479. <https://doi.org/10.3389/fneur.2017.00479>.
56. Budzyński J, Wiśniewska J, Ciecierski M, Kędzia A. Association between bacterial infection and peripheral vascular disease: a review. *Int J Angiol.* 2016;25(1):3–13. <https://doi.org/10.1055/s-0035-1547385>.
57. Fukui Y, Hishikawa N, Shang J, *et al.* Peripheral arterial endothelial dysfunction of neurodegenerative diseases. *J Neurol Sci.* 2016;366:94–9. <https://doi.org/10.1016/j.jns.2016.04.042>.
58. Yamadera M, Fujimura H, Inoue K, *et al.* Microvascular disturbance with decreased pericyte coverage is prominent in the ventral horn of patients with amyotrophic lateral sclerosis. *Amyotroph Lateral Scler Frontotemporal Degener.* 2015;16(5–6):393–401. <https://doi.org/10.3109/21678421.2015.1011663>.
59. Schmid B, Hruscha A, Hogl S, *et al.* Loss of ALS-associated TDP-43 in zebrafish causes muscle degeneration, vascular dysfunction, and reduced motor neuron axon outgrowth. *Proc Natl Acad Sci U S A.* 2013;110(13):4986–91. <https://doi.org/10.1073/pnas.1218311110>.
60. Umahara T, Uchihara T, Hirao K, *et al.* Frontotemporal dementia-associated protein "phosphorylated TDP-43" localizes to atherosclerotic lesions of human carotid and main cerebral arteries. *Histol Histopathol.* 2020;35(2):159–67. <https://doi.org/10.14670/HH-18-140>.
61. McKee AC, Gavett BE, Stern RA, *et al.* TDP-43 proteinopathy and motor neuron disease in chronic traumatic encephalopathy. *J Neuropathol Exp Neurol.* 2010;69(9):918–29. <https://doi.org/10.1097/NEN.0b013e3181ee7d85>.

62. Pupillo E, Poloni M, Bianchi E, *et al.* Trauma and amyotrophic lateral sclerosis: a European population-based case-control study from the EURALS consortium. *Amyotroph Lateral Scler Frontotemporal Degener*. 2018;19(1–2):118–25. <https://doi.org/10.1080/21678421.2017.1386687>.
63. Winkler EA, Sengillo JD, Sullivan JS, *et al.* Blood-spinal cord barrier breakdown and pericyte reductions in amyotrophic lateral sclerosis. *Acta Neuropathol*. 2013;125(1):111–20. <https://doi.org/10.1007/s00401-012-1039-8>.
64. Heyburn L, Abutarboush R, Goodrich S, *et al.* Repeated low-level blast overpressure leads to endovascular disruption and alterations in TDP-43 and Piezo2 in a rat model of blast TBI. *Front Neurol*. 2019;10:766. <https://doi.org/10.3389/fneur.2019.00766>.
65. Aundhakar SC, Mahajan SK, Chhapra DA. Hirayama's disease: a rare clinical variant of amyotrophic lateral sclerosis. *Adv Biomed Res*. 2017;6:95. <https://doi.org/10.4103/2277-9175.211797>.
66. Storti B, Diamanti S, Tremolizzo L, *et al.* ALS mimics due to affection of the cervical spine: from common compressive myelopathy to rare CSF epidural collection. *Case Rep Neurol*. 2021;13(1):145–56. <https://doi.org/10.1159/000512810>.
67. Iliff JJ, Wang M, Liao Y, *et al.* A paravascular pathway facilitates CSF flow through the brain parenchyma and the clearance of interstitial solutes, including amyloid β . *Sci Transl Med*. 2012;4(147):147ra111. <https://doi.org/10.1126/scitranslmed.3003748>.
68. Pappolla M, Sambamurti K, Vidal R, *et al.* Evidence for lymphatic A β clearance in Alzheimer's transgenic mice. *Neurobiol Dis*. 2014;71:215–9. <https://doi.org/10.1016/j.nbd.2014.07.012>.
69. de Leon MJ, Li Y, Okamura N, *et al.* Cerebrospinal fluid clearance in Alzheimer disease measured with dynamic PET. *J Nucl Med*. 2017;58(9):1471–6. <https://doi.org/10.2967/jnumed.116.187211>.
70. Zou W, Pu T, Feng W, *et al.* Blocking meningeal lymphatic drainage aggravates Parkinson's disease-like pathology in mice overexpressing mutated α -synuclein. *Transl Neurodegener*. 2019;8:7. <https://doi.org/10.1186/s40035-019-0147-y>.
71. Sass LR, Khani M, Romm J, *et al.* Non-invasive MRI quantification of cerebrospinal fluid dynamics in amyotrophic lateral sclerosis patients. *Fluids Barriers CNS*. 2020;17(1):4. <https://doi.org/10.1186/s12987-019-0164-3>.
72. Flanagan MF. The role of the craniocervical junction in craniospinal hydrodynamics and neurodegenerative conditions. *Neurol Res Int*. 2015;2015:794829. <https://doi.org/10.1155/2015/794829>.
73. Holzapfel K, Naumann M. Ultrasound detection of vagus nerve atrophy in bulbar amyotrophic lateral sclerosis. *J Neuroimaging*. 2020;30(6):762–5. <https://doi.org/10.1111/jon.12761>.
74. Pathak S, Tripathi S, Deori N, *et al.* Effect of tetracycline family of antibiotics on actin aggregation, resulting in the formation of Hirano bodies responsible for neuropathological disorders. *J Biomol Struct Dyn*. 2021;39(1):236–53. <https://doi.org/10.1080/07391102.2020.1717629>.
75. Altman T, Ionescu A, Ibraheem A, *et al.* Axonal TDP-43 condensates drive neuromuscular junction disruption through inhibition of local synthesis of nuclear encoded mitochondrial proteins. *Nat Commun*. 2021;12(1):6914. <https://doi.org/10.1038/s41467-021-27221-8>.
76. Ng HY, Oliver BG, Burgess JK, *et al.* Doxycycline reduces the migration of tuberous sclerosis complex-2 null cells - effects on RhoA-GTPase and focal adhesion kinase. *J Cell Mol Med*. 2015;19(11):2633–46. <https://doi.org/10.1111/jcmm.12593>.
77. González-Lizárraga F, Socías SB, Ávila CL, *et al.* Repurposing doxycycline for synucleinopathies: remodelling of α -synuclein oligomers towards non-toxic parallel beta-sheet structured species. *Sci Rep*. 2017;7:41755. <https://doi.org/10.1038/srep41755>.
78. Das AT, Tenenbaum L, Berkhout B. Tet-On systems for doxycycline-inducible gene expression. *Curr Gene Ther*. 2016;16(3):156–67. <https://doi.org/10.2174/156652321666160524144041>.
79. Gasparrini AJ, Markley JL, Kumar H, *et al.* Tetracycline-inactivating enzymes from environmental, human commensal, and pathogenic bacteria cause broad-spectrum tetracycline resistance. *Commun Biol*. 2020;3(1):241. <https://doi.org/10.1038/s42003-020-0966-5>.
80. Stone LK, Baym M, Lieberman TD, *et al.* Compounds that select against the tetracycline-resistance efflux pump. *Nat Chem Biol*. 2016;12(11):902–4. <https://doi.org/10.1038/nchembio.2176>.
81. Domon H, Hiyoshi T, Maekawa T, *et al.* Antibacterial activity of hinokitiol against both antibiotic-resistant and -susceptible pathogenic bacteria that predominate in the oral cavity and upper airways. *Microbiol Immunol*. 2019;63(6):213–22. <https://doi.org/10.1111/1348-0421.12688>.
82. Kim DJ, Lee MW, Choi JS, *et al.* Inhibitory activity of hinokitiol against biofilm formation in fluconazole-resistant *Candida* species. *PLoS One*. 2017;12(2):e0171244. <https://doi.org/10.1371/journal.pone.0171244>.
83. Lee JH, Jeong JK, Park SY. AMPK Activation mediated by hinokitiol inhibits adipogenic differentiation of mesenchymal stem cells through autophagy flux. *Int J Endocrinol*. 2018;2018:2014192. <https://doi.org/10.1155/2018/2014192>.
84. Moon JH, Lee JH, Lee YJ, Park SY. Hinokitiol protects primary neuron cells against prion peptide-induced toxicity via autophagy flux regulated by hypoxia inducing factor-1. *Oncotarget*. 2016;7(21):29944–57. <https://doi.org/10.18632/oncotarget.8670>.
85. Grillo AS, SantaMaria AM, Kafina MD, *et al.* Restored iron transport by a small molecule promotes absorption and hemoglobinization in animals. *Science*. 2017;356(6338):608–16. <https://doi.org/10.1126/science.aah3862>.

86. Zhou J, Lu Y, Wu W, Feng Y. Taurine promotes the production of CD4+CD25+FOXP3+ Treg cells through regulating IL-35/STAT1 pathway in a mouse allergic rhinitis model. *Allergy Asthma Clin Immunol.* 2021;17(1):59. <https://doi.org/10.1186/s13223-021-00562-1>.
87. Kato J, Ido A, Hasuike S, *et al.* Transforming growth factor-beta-induced stimulation of formation of collagen fiber network and anti-fibrotic effect of taurine in an in vitro model of hepatic fibrosis. *Hepatology Res.* 2004;30(1):34–41. <https://doi.org/10.1016/j.hepres.2004.04.006>.
88. Jung MK, Kim KY, Lee NY, *et al.* Expression of taurine transporter (TauT) is modulated by heat shock factor 1 (HSF1) in motor neurons of ALS. *Mol Neurobiol.* 2013;47(2):699–710. <https://doi.org/10.1007/s12035-012-8371-9>.
89. Kim W, Kim HU, Lee HN, *et al.* Taurine chloramine stimulates efferocytosis through upregulation of Nrf2-mediated heme oxygenase-1 expression in murine macrophages: possible involvement of carbon monoxide. *Antioxid Redox Signal.* 2015;23(2):163–77. <https://doi.org/10.1089/ars.2013.5825>.
90. Boussicault L, Laffaire J, Schmitt P, *et al.* Combination of acamprosate and baclofen (PXT864) as a potential new therapy for amyotrophic lateral sclerosis. *J Neurosci Res.* 2020;98(12):2435–50. <https://doi.org/10.1002/jnr.24714>.
91. Kalk NJ, Lingford-Hughes AR. The clinical pharmacology of acamprosate. *Br J Clin Pharmacol.* 2014;77(2):315–23. <https://doi.org/10.1111/bcp.12070>.
92. Lienhart WD, Gudipati V, Macheroux P. The human flavoproteome. *Arch Biochem Biophys.* 2013;535(2):150162. <https://doi.org/10.1016/j.abb.2013.02.015>.
93. Lei J, Xin C, Xiao W, *et al.* The promise of endogenous and exogenous riboflavin in antiinfection. *Virulence.* 2021;12(1):2314–26. <https://doi.org/10.1080/21505594.2021.1963909>.
94. Depeint F, Bruce WR, Shangari N, *et al.* Mitochondrial function and toxicity: role of the B vitamin family on mitochondrial energy metabolism. *Chem Biol Interact.* 2006;163(1-2):94–112. <https://doi.org/10.1016/j.cbi.2006.04.014>.
95. Parikh S, Goldstein A, Koenig MK, *et al.* Diagnosis and management of mitochondrial disease: a consensus statement from the Mitochondrial Medicine Society. *Genet Med.* 2015;17(9):689–701. <https://doi.org/10.1038/gim.2014.177>.
96. Rizzo F, Ramirez A, Compagnucci C, *et al.* Genome-wide RNA-seq of iPSC-derived motor neurons indicates selective cytoskeletal perturbation in Brown-Vialetto disease that is partially rescued by riboflavin. *Sci Rep.* 2017;7:46271. <https://doi.org/10.1038/srep46271>.
97. Carreau C, Lenglet T, Mosnier I, *et al.* A juvenile ALS-like phenotype dramatically improved after high-dose riboflavin treatment. *Ann Clin Transl Neurol.* 2020;7(2):250–3. <https://doi.org/10.1002/acn3.50977>.
98. Toyosawa T, Suzuki M, Kodama K, Araki S. Effects of intravenous infusion of highly purified vitamin B2 on lipopolysaccharide-induced shock and bacterial infection in mice. *Eur J Pharmacol.* 2004;492(2-3):273–80. <https://doi.org/10.1016/j.ejphar.2004.04.004>.
99. Lee ER, Blount KF, Breaker RR. Roseoflavin is a natural antibacterial compound that binds to FMN riboswitches and regulates gene expression. *RNA Biol.* 2009;6(2):187–94. <https://doi.org/10.4161/rna.6.2.7727>.
100. Scafidi S, Fiskum G, Lindauer SL, *et al.* Metabolism of acetyl-L-carnitine for energy and neurotransmitter synthesis in the immature rat brain. *J Neurochem.* 2010;114(3):820–31. <https://doi.org/10.1111/j.1471-4159.2010.06807.x>.
101. Borghi-Silva A, Baldissera V, Sampaio LM, *et al.* L-carnitine as an ergogenic aid for patients with chronic obstructive pulmonary disease submitted to whole-body and respiratory muscle training programs. *Braz J Med Biol Res.* 2006;39(4):465–74. <https://doi.org/10.1590/s0100-879x2006000400006>.
102. Bavbek N, Akay H, Uz B, *et al.* The effects of L-carnitine therapy on respiratory function tests in chronic hemodialysis patients. *Ren Fail.* 2010;32(2):157–61. <https://doi.org/10.3109/08860221003592812>.
103. Ogle RF, Christodoulou J, Fagan E, *et al.* Mitochondrial myopathy with tRNA(Leu(UUR)) mutation and complex I deficiency responsive to riboflavin. *J Pediatr.* 1997;130(1):138–45. [https://doi.org/10.1016/s00223476\(97\)70323-8](https://doi.org/10.1016/s00223476(97)70323-8).
104. Shimokawa H, Sunamura S, Satoh K. RhoA/Rho-kinase in the cardiovascular system. *Circ Res.* 2016;118(2):352–66. <https://doi.org/10.1161/CIRCRESAHA.115.306532>.
105. Masumoto A, Mohri M, Shimokawa H, *et al.* Suppression of coronary artery spasm by the Rho-kinase inhibitor fasudil in patients with vasospastic angina. *Circulation.* 2002;105(13):1545–7. <https://doi.org/10.1161/hc1002.105938>.
106. Olson MF. Applications for ROCK kinase inhibition. *Curr Opin Cell Biol.* 2008;20(2):242–8. <https://doi.org/10.1016/j.ccb.2008.01.002>.
107. Suzuki Y, Shibuya M, Satoh S, *et al.* Safety and efficacy of fasudil monotherapy and fasudil-ozagrel combination therapy in patients with subarachnoid hemorrhage: sub-analysis of the postmarketing surveillance study. *Neurol Med Chir (Tokyo).* 2008;48(6):241–8. <https://doi.org/10.2176/nmc.48.241>.
108. Velat GJ, Kimball MM, Mocco JD, Hoh BL. Vasospasm after aneurysmal subarachnoid hemorrhage: review of randomized controlled trials and meta-analyses in the literature. *World Neurosurg.* 2011;76(5):446–54. <https://doi.org/10.1016/j.wneu.2011.02.030>.
109. Rao PV, Pattabiraman PP, Kocczynski C. Role of the Rho GTPase/Rho kinase signaling pathway in pathogenesis and treatment of glaucoma: bench to bedside research. *Exp Eye Res.* 2017;158:23–32. <https://doi.org/10.1016/j.exer.2016.08.023>.

110. Stankiewicz TR, Ramaswami SA, Bouchard RJ, *et al.* Neuronal apoptosis induced by selective inhibition of Rac GTPase versus global suppression of Rho family GTPases is mediated by alterations in distinct mitogen-activated protein kinase signaling cascades. *J Biol Chem.* 2015;290(15):9363–76. <https://doi.org/10.1074/jbc.M114.575217>.
111. Markovinic A, Cimbro R, Ljutic T, *et al.* Optineurin in amyotrophic lateral sclerosis: Multifunctional adaptor protein at the crossroads of different neuroprotective mechanisms. *Prog Neurobiol.* 2017;154:1–20. <https://doi.org/10.1016/j.pneurobio.2017.04.005>.
112. Slowicka K, Vereecke L, van Loo G. Cellular functions of optineurin in health and disease. *Trends Immunol.* 2016;37(9):621–33. <https://doi.org/10.1016/j.it.2016.07.002>.
113. Toth RP, Atkin JD. Dysfunction of optineurin in amyotrophic lateral sclerosis and glaucoma. *Front Immunol.* 2018;9:1017. <https://doi.org/10.3389/fimmu.2018.01017>.
114. Lingor P, Weber M, Camu W, *et al.* ROCK-ALS: Protocol for a randomized, placebo-controlled, double-blind phase IIa trial of safety, tolerability and efficacy of the Rho kinase (ROCK) inhibitor fasudil in amyotrophic lateral sclerosis. *Front Neurol.* 2019;10:293. <https://doi.org/10.3389/fneur.2019.00293>.
115. Kulkarni SK, Dhir A. sigma-1 receptors in major depression and anxiety. *Expert Rev Neurother.* 2009;9(7):1021–34. <https://doi.org/10.1586/ern.09.40>.
116. Huang L, Zhang X, Ma X, *et al.* Berberine alleviates endothelial glycocalyx degradation and promotes glycocalyx restoration in LPS-induced ARDS. *Int Immunopharmacol.* 2018;65:96–107. <https://doi.org/10.1016/j.intimp.2018.10.001>.
117. Jin Y, Liu S, Ma Q, *et al.* Berberine enhances the AMPK activation and autophagy and mitigates high glucose-induced apoptosis of mouse podocytes. *Eur J Pharmacol.* 2017;794:106–14. <https://doi.org/10.1016/j.ejphar.2016.11.037>.
118. Chang CF, Lee YC, Lee KH, *et al.* Therapeutic effect of berberine on TDP-43-related pathogenesis in FTLN and ALS. *J Biomed Sci.* 2016;23(1):72. <https://doi.org/10.1186/s12929-016-0290-z>.
119. Rusmini P, Cristofani R, Tedesco B, *et al.* Enhanced clearance of neurotoxic misfolded proteins by the natural compound berberine and its derivatives. *Int J Mol Sci.* 2020;21(10):3443. <https://doi.org/10.3390/ijms21103443>.
120. Zeng P, Zhou X. Causal effects of blood lipids on amyotrophic lateral sclerosis: a Mendelian randomization study. *Hum Mol Genet.* 2019;28(4):688–97. <https://doi.org/10.1093/hmg/ddy384>.
121. Hope J. A review of the mechanism of injury and treatment approaches for illness resulting from exposure to water-damaged buildings, mold, and mycotoxins. *ScientificWorldJournal.* 2013;2013:767482. <https://doi.org/10.1155/2013/767482>.
122. Rezaei AR, Ranjan M, D'Haese PF, *et al.* Noninvasive hippocampal blood-brain barrier opening in Alzheimer's disease with focused ultrasound. *Proc Natl Acad Sci U S A.* 2020;117(17):9180–2. <https://doi.org/10.1073/pnas.2002571117>.
123. Yu FTH, Chen X, Straub AC, Pacella JJ. The role of nitric oxide during sonoreperfusion of microvascular obstruction. *Theranostics.* 2017;7(14):3527–38. <https://doi.org/10.7150/thno.19422>.
124. Beggs CB. Venous hemodynamics in neurological disorders: an analytical review with hydrodynamic analysis. *BMC Med.* 2013;11:142. <https://doi.org/10.1186/1741-7015-11-142>.
125. Nichols NL, Van Dyke J, Nashold L, *et al.* Ventilatory control in ALS. *Respir Physiol Neurobiol.* 2013;189(2):429–437. <https://doi.org/10.1016/j.resp.2013.05.016>.
126. de Carvalho M, Swash M, Pinto S. Diaphragmatic neurophysiology and respiratory markers in ALS. *Front Neurol.* 2019;10:143. <https://doi.org/10.3389/fneur.2019.00143>.
127. Niedermeyer S, Murn M, Choi PJ. Respiratory failure in amyotrophic lateral sclerosis. *Chest.* 2019;155(2):401–8. <https://doi.org/10.1016/j.chest.2018.06.035>.
128. Puls I, Beck M, Giess R, *et al.* Clenbuterol bei der Amyotrophen Lateralsklerose. Kein Hinweis für einen positiven Effekt [Clenbuterol in amyotrophic lateral sclerosis. No indication for a positive effect]. *Nervenarzt.* 1999;70(12):1112–5. <https://doi.org/10.1007/s001150050548>.
129. Zafar A, Zulfikar H. Aminophylline. [Updated 2022 Jan 7]. In: StatPearls [Internet]. Treasure Island (FL): StatPearls Publishing; 2022 Jan-. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK545175/>.
130. Golder FJ, Ranganathan L, Satriotomo I, *et al.* Spinal adenosine A2a receptor activation elicits long-lasting phrenic motor facilitation. *J Neurosci.* 2008;28(9):2033–42. <https://doi.org/10.1523/JNEUROSCI.3570-07.2008>.
131. Berto MC, Filha SC, Camelier A, *et al.* Acute action of aminophylline in patients with amyotrophic lateral sclerosis. *Acta Neurol Scand.* 2007;115(5):301–5. <https://doi.org/10.1111/j.1600-0404.2007.00643.x>.
132. Massey EW, Lowe S. Lithium carbonate in pseudobulbar palsy. *Ann Neurol.* 1981;9(1):97. <https://doi.org/10.1002/ana.410090128>.
133. Liu M, Qian T, Zhou W, *et al.* Beneficial effects of low-dose lithium on cognitive ability and pathological alteration of Alzheimer's disease transgenic mice model. *Neuroreport.* 2020;31(13):943–51. <https://doi.org/10.1097/WNR.0000000000001499>.
134. Moutal A, White KA, Chefdeville A, *et al.* Dysregulation of CRMP2 post-translational modifications drive its pathological functions. *Mol Neurobiol.* 2019;56(10):6736–55. <https://doi.org/10.1007/s12035-019-1568-4>.

135. Lin PC, Chan PM, Hall C, Manser E. Collapsin response mediator proteins (CRMPs) are a new class of microtubule-associated protein (MAP) that selectively interacts with assembled microtubules via a taxol-sensitive binding interaction. *J Biol Chem*. 2011;286(48):41466–78. <https://doi.org/10.1074/jbc.M111.283580>.
136. Nakamura F, Ohshima T, Goshima Y. Collapsin response mediator proteins: their biological functions and pathophysiology in neuronal development and regeneration. *Front Cell Neurosci*. 2020;14:188. <https://doi.org/10.3389/fncel.2020.00188>.
137. Brot S, Auger C, Bentata R, et al. Collapsin response mediator protein 5 (CRMP5) induces mitophagy, thereby regulating mitochondrion numbers in dendrites. *J Biol Chem*. 2014;289(4):2261–76. <https://doi.org/10.1074/jbc.M113.490862>.
138. Alabed YZ, Pool M, Ong Tone S, Fournier AE. Identification of CRMP4 as a convergent regulator of axon outgrowth inhibition. *J Neurosci*. 2007;27(7):1702–11. <https://doi.org/10.1523/JNEUROSCI.5055-06.2007>.
139. Numata-Uematsu Y, Wakatsuki S, Nagano S, et al. Inhibition of collapsin response mediator protein-2 phosphorylation ameliorates motor phenotype of ALS model mice expressing SOD1^{G93A}. *Neurosci Res*. 2019;139:63–8. <https://doi.org/10.1016/j.neures.2018.08.016>.
140. Arimura N, Ménager C, Kawano Y, et al. Phosphorylation by Rho kinase regulates CRMP-2 activity in growth cones. *Mol Cell Biol*. 2005;25(22):9973–84. <https://doi.org/10.1128/MCB.25.22.9973-9984.2005>.
141. Yang W, Leystra-Lantz C, Strong MJ. Upregulation of GSK3beta expression in frontal and temporal cortex in ALS with cognitive impairment (ALS_{ci}). *Brain Res*. 2008;1196:131–9. <https://doi.org/10.1016/j.brainres.2007.12.031>.
142. He Z, Zhou Y, Wang Q, et al. Inhibiting endoplasmic reticulum stress by lithium chloride contributes to the integrity of blood-spinal cord barrier and functional recovery after spinal cord injury. *Am J Transl Res*. 2017;9(3):1012–24. PMID: 28386329.
143. Tong M, He Z, Lin X, et al. Lithium chloride contributes to blood-spinal cord barrier integrity and functional recovery from spinal cord injury by stimulating autophagic flux. *Biochem Biophys Res Commun*. 2018;495(4):2525–31. <https://doi.org/10.1016/j.bbrc.2017.12.119>.
144. Fornai F, Longone P, Cafaro L, et al. Lithium delays progression of amyotrophic lateral sclerosis [published correction appears in *Proc Natl Acad Sci U S A*. 2008 21;105(42):16404–7]. *Proc Natl Acad Sci U S A*. 2008;105(6):2052–7. <https://doi.org/10.1073/pnas.0708022105>.
145. Pasquali L, Longone P, Isidoro C, et al. Autophagy, lithium, and amyotrophic lateral sclerosis. *Muscle Nerve*. 2009;40(2):173–94. <https://doi.org/10.1002/mus.21423>.
146. Devanand DP, Pelton GH, D'Antonio K, et al. Low-dose lithium treatment for agitation and psychosis in Alzheimer disease and frontotemporal dementia: a case series. *Alzheimer Dis Assoc Disord*. 2017;31(1):73–5. <https://doi.org/10.1097/WAD.0000000000000161>.
147. van Eijk RPA, Jones AR, Sproviero W, et al. Meta-analysis of pharmacogenetic interactions in amyotrophic lateral sclerosis clinical trials [published correction appears in *Neurology*. 2017;89(22):2303]. *Neurology*. 2017;89(18):1915–22. <https://doi.org/10.1212/WNL.0000000000004606>.
148. Limanaqi F, Biagioni F, Ryskalin L, et al. Molecular mechanisms linking ALS/FTD and psychiatric disorders, the potential effects of lithium. *Front Cell Neurosci*. 2019;13:450. <https://doi.org/10.3389/fncel.2019.00450>.
149. Ionescu A, Gradus T, Altman T, et al. Targeting the sigma-1 receptor via pridopidine ameliorates central features of ALS pathology in a SOD1^{G93A} model. *Cell Death Dis*. 2019;10(3):210. <https://doi.org/10.1038/s41419-019-1451-2>.
150. Mancuso R, Oliván S, Rando A, et al. Sigma-1R agonist improves motor function and motoneuron survival in ALS mice. *Neurotherapeutics*. 2012;9(4):814–26. <https://doi.org/10.1007/s13311-012-0140-y>.
151. Ono Y, Tanaka H, Takata M, et al. SA4503, a sigma-1 receptor agonist, suppresses motor neuron damage in in vitro and in vivo amyotrophic lateral sclerosis models. *Neurosci Lett*. 2014;559:174–8. <https://doi.org/10.1016/j.neulet.2013.12.005>.
152. Mavlyutov TA, Guo LW, Epstein ML, Ruoho AE. Role of the sigma-1 receptor in amyotrophic lateral sclerosis (ALS). *J Pharmacol Sci*. 2015;127(1):10–6. <https://doi.org/10.1016/j.jphs.2014.12.013>.
153. Liu DY, Chi TY, Ji XF, et al. Sigma-1 receptor activation alleviates blood-brain barrier dysfunction in vascular dementia mice. *Exp Neurol*. 2018;308:90–9. <https://doi.org/10.1016/j.expneurol.2018.07.002>.
154. Vandoorne T, De Bock K, Van Den Bosch L. Energy metabolism in ALS: an underappreciated opportunity? *Acta Neuropathol*. 2018;135(4):489–509. <https://doi.org/10.1007/s00401-018-1835-x>.
155. Patel BP, Safdar A, Raha S, et al. Caloric restriction shortens lifespan through an increase in lipid peroxidation, inflammation and apoptosis in the G93A mouse, an animal model of ALS. *PLoS One*. 2010;5(2):e9386. <https://doi.org/10.1371/journal.pone.0009386>.
156. Coughlan KS, Halang L, Woods I, Prehn JH. A high-fat jelly diet restores bioenergetic balance and extends lifespan in the presence of motor dysfunction and lumbar spinal cord motor neuron loss in TDP-43A315T mutant C57BL/6J mice. *Dis Model Mech*. 2016;9(9):1029–37. <https://doi.org/10.1242/dmm.024786>.
157. Dupuis L, Oudart H, René F, et al. Evidence for defective energy homeostasis in amyotrophic lateral sclerosis: benefit of a high-energy diet in a transgenic mouse model. *Proc Natl Acad Sci U S A*. 2004;101(30):11159–64. <https://doi.org/10.1073/pnas.0402026101>.

158. Wills AM, Hubbard J, Macklin EA, *et al.* Hypercaloric enteral nutrition in patients with amyotrophic lateral sclerosis: a randomised, double-blind, placebo-controlled phase 2 trial. *Lancet*. 2014;383(9934):2065–72. [https://doi.org/10.1016/S0140-736\(14\)60222-1](https://doi.org/10.1016/S0140-736(14)60222-1).
159. Calignano A, La Rana G, Piomelli D. Antinociceptive activity of the endogenous fatty acid amide, palmitylethanolamide. *Eur J Pharmacol*. 2001;419(2–3):191–8. [https://doi.org/10.1016/s0014-2999\(01\)00988-8](https://doi.org/10.1016/s0014-2999(01)00988-8).
160. Sabelli H, Fink P, Fawcett J, Tom C. Sustained antidepressant effect of PEA replacement. *J Neuropsychiatry Clin Neurosci*. 1996;8(2):168–71. <https://doi.org/10.1176/jnp.8.2.168>.
161. Lin TY, Lu CW, Wu CC, *et al.* Palmitoylethanolamide inhibits glutamate release in rat cerebrocortical nerve terminals. *Int J Mol Sci*. 2015;16(3):5555–71. <https://doi.org/10.3390/ijms16035555>.
162. Palma E, Reyes-Ruiz JM, Loperogo D, *et al.* Acetylcholine receptors from human muscle as pharmacological targets for ALS therapy. *Proc Natl Acad Sci U S A*. 2016;113(11):3060–5. <https://doi.org/10.1073/pnas.1600251113>.
163. Giannoni A, Borrelli C, Mirizzi G, *et al.* Benefit of buspirone on chemoreflex and central apnoeas in heart failure: a randomized controlled crossover trial. *Eur J Heart Fail*. 2021;23(2):312320. <https://doi.org/10.1002/ejhf.1854>.
164. Maresh S, Prowting J, Vaughan S, *et al.* Buspirone decreases susceptibility to hypocapnic central sleep apnea in chronic SCI patients. *J Appl Physiol (1985)*. 2020;129(4):675–82. <https://doi.org/10.1152/jappphysiol.00435.2020>.
165. Vollrath JT, Sechi A, Dreser A, *et al.* Loss of function of the ALS protein SigR1 leads to ER pathology associated with defective autophagy and lipid raft disturbances. *Cell Death Dis*. 2014;5(6):e1290. <https://doi.org/10.1038/cddis.2014.243>.
166. Meininger V, Bensimon G, Bradley WR, *et al.* Efficacy and safety of xaliproden in amyotrophic lateral sclerosis: results of two phase III trials. *Amyotroph Lateral Scler Other Motor Neuron Disord*. 2004;5(2):107117. <https://doi.org/10.1080/14660820410019602>.
167. Langade D, Thakare V, Kanchi S, Kelgane S. Clinical evaluation of the pharmacological impact of ashwagandha root extract on sleep in healthy volunteers and insomnia patients: A double-blind, randomized, parallel-group, placebo-controlled study. *J Ethnopharmacol*. 2021;264:113276. <https://doi.org/10.1016/j.jep.2020.113276>.
168. Bale S, Venkatesh P, Sunkoju M, Godugu C. An adaptogen: withaferin A ameliorates *in vitro* and *in vivo* pulmonary fibrosis by modulating the interplay of fibrotic, matricellular proteins, and cytokines. *Front Pharmacol*. 2018;9:248. <https://doi.org/10.3389/fphar.2018.00248>.
169. Behl T, Sharma A, Sharma L, *et al.* Exploring the multifaceted therapeutic potential of withaferin A and its derivatives. *Biomedicines*. 2020;8(12):571. <https://doi.org/10.3390/biomedicines8120571>.
170. Kumar S, Phaneuf D, Cordeau P Jr, *et al.* Induction of autophagy mitigates TDP-43 pathology and translational repression of neurofilament mRNAs in mouse models of ALS/FTD. *Mol Neurodegener*. 2021;16(1):1. <https://doi.org/10.1186/s13024-020-00420-5>.
171. Pinato L, da Silveira Cruz-Machado S, Franco DG, *et al.* Selective protection of the cerebellum against intracerebroventricular LPS is mediated by local melatonin synthesis. *Brain Struct Funct*. 2015;220(2):827–40. <https://doi.org/10.1007/s00429-013-0686-4>.
172. Hu Y, Wang Z, Pan S, *et al.* Melatonin protects against blood-brain barrier damage by inhibiting the TLR4 NFκB signaling pathway after LPS treatment in neonatal rats. *Oncotarget*. 2017;8(19):31638–54. <https://doi.org/10.18632/oncotarget.15780>.
173. Sagrillo-Fagundes L, Bienvenue-Pariseault J, Vaillancourt C. Melatonin: the smart molecule that differentially modulates autophagy in tumor and normal placental cells. *PLoS One*. 2019;14(1):e0202458. <https://doi.org/10.1371/journal.pone.0202458>.
174. Bald EM, Nance CS, Schultz JL. Melatonin may slow disease progression in amyotrophic lateral sclerosis: findings from the pooled resource open-access ALS clinic trials database. *Muscle Nerve*. 2021;63(4):572576. <https://doi.org/10.1002/mus.27168>.